# Synthesis of Analogues of Brassinosteroids from Chenodeoxycholic Acid

Roxana Pérez Gil<sup>a</sup>, Martín Andrés Iglesias Arteaga<sup>a</sup>, Carlos Pérez Martínez<sup>a</sup>, Francisco Coll Manchado<sup>\*a</sup>, Danahe Coll García<sup>a</sup>, and Arístides Rosado<sup>b</sup>

Laboratorio de Productos Naturales de la Facultad de Química, Universidad de la Habana<sup>a</sup>,

Zapata y G, Ciudad de La Habana, Cuba 10400

E-mail: roxana@karin.fmq.uh.edu.cu

Centro Nacional de Investigaciones Cientificas<sup>b</sup>, Avenida 25 y calle 128, Ciudad de La Habana, Cuba

Received January 27, 1998

Keywords: Natural products / Steroids / Chenodeoxycholic acid / Brassinosteroid analogues / Biological activity

The synthesis and spectroscopic characterization of two new bioactive analogues of brassinosteroids with a 24hydroxylated cholanic side chain, an A/B ring *cis*-junction and oxygenated functions in C-7 is described.

#### Introduction

Brassinosteroids are plant hormones<sup>[1]</sup> with remarkable biological activity that have stimulated many laboratories to undertake their synthesis<sup>[2]</sup> and the study of the relationship between molecular structure and biological activity. <sup>[3]</sup> Although it is generally accepted that a high brassinosteroid activity requires (a) a  $2\alpha$ , $3\alpha$ -diol in the A ring, (b) a 6-one or 6-oxo-7-oxalactone moiety in the B ring, (c) an A/B *trans*-fused ring junction, (d) a (22R,23R)-diol and (e) a 24-methyl or -ethyl substituent, <sup>[1]</sup> a great variety of bioactive analogues with modifications in the steroidal nucleus and side chain have been synthesized. <sup>[4]</sup> Recently, we have reported on the synthesis of some bioactive spirostanic <sup>[5]</sup> and

furostanic <sup>[6]</sup> analogues of brassinosteroids starting from steroidal sapogenins. We now wish to describe the synthesis of two new bioactive analogues of brassinosteroids with a 24-hydroxylated cholanic side chain, an A/B ring *cis* junction and oxygenated functions in C-7.

#### **Results and Discussion**

Methyl chenodeoxycholate (2) was obtained by a procedure already described. <sup>[7]</sup> LiAlH<sub>4</sub> reduction of 2 in tetrahydrofuran (THF) led to the triol  $3^{[8]}$  which was selectively cathylated to afford the  $7\alpha$ -hydroxy- $5\beta$ -cholan- $3\alpha$ ,24-diyl dicathylate (4). Oxidation of 4, followed by hydrolysis with 5% methanolic KOH afforded  $3\alpha$ ,24-dihydroxy- $5\beta$ -cholan-

Scheme 1. Synthetic sequence

F. Coll Manchado et al.

7-one (6) in 79% overall yield. Baeyer-Villiger reaction of 5 with trifluoroperoxyacetic acid (TFPAA) obtained "in situ" by the trifluoroacetic anhydride (TFAA)/hydrogen peroxide procedure, <sup>[8]</sup> led to pure lactone 7. Selective hydrolysis of 7 with 1% methanolic  $K_2CO_3$  during 15 hours afforded the desired compound  $3\alpha,24$ -dihydroxy-B-homo-8-oxa-5 $\beta$ -cholan-7-one (8) in 66% overall yield (Scheme 1).

The biological activity of the brassinosteroid analogues **6** and **8** was examined by the Radish test. <sup>[9]</sup> Compound **6** produced, at the concentration of  $10^{-6}$  µg/ml, an increase of 36% (referred to an untreated control) in the weight of cotyledons and at  $10^{-4}$  and  $10^{-6}$  µg/ml an increase of 16 and 17% respectively, in the length of hypocothyls. Compound **8** produced at the concentration of  $10^{-4}$  µg/ml just a small increase of 9% in the weight of cotyledons.

### **Conclusions**

Two new analogues of brassinosteroids **6** and **8** with an A/B ring *cis* junction, oxygenated functions in C-7 and a 24-hydroxylated cholanic side chain were synthesized from chenodeoxycholic acid (1). In contrast with the expected results, the compounds which bear drastic structural modifications, showed biological activity. The biological activity of the 7-oxo compound **6** proved to be higher than that of the lactone **8**.

## **Experimental Section**

General: IR spectra were recorded from KBr cells with a Phillips Analytical PU9800 FT-IR spectrometer. — NMR: Bruker 250 ACF (250 MHz and 62.9 MHz, for  $^1H$  and  $^{13}C$ , respectively). For  $^1H$  NMR, CDCl $_3$  or CD $_3$ OD (solvents) and TMS (internal standard); for  $^{13}C$  NMR, CDCl $_3$  or CD $_3$ OD ( $\delta_C=77.0$  and 49.0) were used. Temperature of the samples was 300 K and concentrations were 30 mg/ml for  $^{13}C$  spectra and 10 mg/ml for  $^1H$  spectra. — MS: JEOL JMS-DX 300 (70 eV). — Melting points were measured with Electrothermal 9100 equipment and are uncorrected.

 $5\beta$ -Cholane- $3\alpha$ ,  $7\alpha$ , 24-triol (3): To a suspension of 500 mg of LiAlH<sub>4</sub> in 15 ml of THF, a solution of 2.54 g (6.72·10<sup>-3</sup> mol) of methyl chenodeoxycholanate (2) in 15 ml of THF was added dropwise, and the mixture was stirred at reflux temp. for 4 h, poured into cool water and 10% H<sub>2</sub>SO<sub>4</sub> (10 ml) was added. The product was extracted with ethyl acetate (3 imes 30 ml) and the combined organic extracts were washed with brine (2 imes 20 ml) and water (2  $\times$  20 ml), dried with MgSO<sub>4</sub> and concentrated to give 2.25 g of 3 (95%), colorless crystals, m.p. 104.8-105.8 °C. – IR (KBr):  $\tilde{v}=$ 3404 cm $^{-1}$  (OH). - <sup>1</sup>H NMR (CD<sub>3</sub>OD):  $\delta = 3.51$  (m, 1 H, 3-H<sub>ax</sub>), 3.79 (m, 1 H, 7-H<sub>eq</sub>), 0.70 (s, 3 H, 18-H), 0.93 (s, 3 H, 19-H), 0.97 (d, J = 6.21 Hz, 3 H, 21-H), 3.40 (t, 2 H, 24-H).  $- {}^{13}$ C NMR (CD<sub>3</sub>OD):  $\delta = 36.9$  (C-1), 33.6 (C-2), 73.2 (C-3), 41.5 (C-4), 43.6 (C-5), 36.3 (C-6), 69.4 (C-7), 41.2 (C-8), 34.4 (C-9), 36.6 (C-10), 22.2 (C-11), 40.8 (C-12), 44.0 (C-13), 51.9 (C-14), 25.1 (C-15), 29.8 (C-16), 57.9 (C-17), 12.7 (C-18), 23.9 (C-19), 37.5 (C-20), 19.7 (C-21), 31.8 (C-22), 30.7 (C-23), 63.9 (C-24). - MS (70 eV); m/z (%): 378 (4.9) [M $^+$ ], 360 (33.6) [M $^+$  -  $H_2O$ ], 342 (71.3) [M $^+$  -  $2H_2O$ ], 283 (2.8) [M $^+$  - 2 H $_2$ O - C $_3$ H $_7$ O], 255 (28.7) [M $^+$  - 2 H $_2$ O - $C_5H_{11}O$ ], 41 (100) [C=C-C<sup>+</sup>]. -  $C_{24}H_{42}O_3$  (378.59): calcd. C 76.14, H 11.18; found C 76.10, H 11.24.

 $7\alpha$ -Hydroxy-5 $\beta$ -cholane-3 $\alpha$ ,24-diyl Dicathylate (4): To a solution of 1.65 g (4.36·10<sup>-3</sup> mol) of 3 in 15 ml of dioxane and 8 ml of

pyridine at 0°C, 10 ml of ethyl chloroformate was added dropwise, and the mixture was stirred at room temp. for 2 h. 8 ml of 32% HCl and 50 ml of water were added and the product was extracted with dichloromethane. The organic phase was washed with water, dried with MgSO<sub>4</sub> and the solvent removed under reduced pressure to give 2.21 g of 4 (97%), white crystalline solid, m.p. 107.4-109.4 °C. – IR (KBr):  $\tilde{v} = 3536$  cm<sup>-1</sup> (OH), 1740 (C=O).  $^{-1}$ H NMR (CDCl<sub>3</sub>):  $\delta = 4.44$  (m, 1 H, 3-H<sub>ax</sub>), 3.84 (m, 1 H, 7- $H_{eq}$ ), 0.66 (s, 3 H, 18-H), 0.92 (s, 3 H, 19-H), 0.93 (d, J = 7.0 Hz, 3 H, 21-H), 4.09 (t, 2 H, 24-H), 1.29-1.31 (t, 6 H, J=7.1 Hz,  $CH_3CH_2OCO_2$ ), 4.16-4.19 (q, J = 7.2 Hz and J = 7.1 Hz, 4 H,  $CH_3CH_2OCO_2$ ). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 34.9$  (C-1), 26.6 (C-2), 78.1 (C-3), 35.1 (C-4), 41.1 (C-5), 34.3 (C-6), 68.3 (C-7), 39.4 (C-7) 8), 32.7 (C-9), 35.0 (C-10), 20.5 (C-11), 39.6 (C-12), 42.6 (C-13), 50.3 (C-14), 23.6 (C-15), 28.2 (C-16), 55.8 (C-17), 11.7 (C-18), 22.6 (C-19), 35.4 (C-20), 18.4 (C-21), 31.7 (C-22), 25.2 (C-23), 68.4 (C-24). 14.3 double intensity  $(CH_3-CH_2-O)$ , 63.5/63.8  $(CH_3 - CH_2 - O)$ , 155.3/154.7  $(O_2C = O)$ .

 $3\alpha$ , 24-Dicathyloxy-5 $\beta$ -cholan-7-one (5): To a solution of 2 g of 4 (3.85·10<sup>-3</sup> mol) in 15 ml of acetone at 0°C, 1.5 ml of Jones reagent was added dropwise and the mixture was stirred for 10 min. 0.5 ml of 2-propanol and enough water to produce a precipitate was added. The precipitate was collected by filtration, washed with abundant water and dried at 60°C to give 1.85 g of 5 (98%), syrup. -IR (KBr):  $\tilde{v} = 1744$ , 1715 cm<sup>-1</sup> (C=O). – <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta =$ 4.54 (m, 1 H, 3-H<sub>ax</sub>), 0.65 (s, 3 H, 18-H), 1.21 (s, 3 H, 19-H), 0.93 (d, J = 6.4 Hz, 3 H, 21-H), 4.09 (t, 2 H, 24-H) 1.29-1.31 (t, J =7.1 Hz, 6 H,  $CH_3CH_2OCO_2$ ), 4.16-4.19 (q, J = 7.1 Hz, 4 H,  $CH_3CH_2OCO_2$ ). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 33.7$  (C-1), 26.0 (C-2), 76.5 (C-3), 33.0 (C-4), 42.6 (C-5), 45.2 (C-6), 211.5 (C-7), 48.9 (C-8), 45.7 (C-9), 35.1 (C-10), 21.7 (C-11), 38.9 (C-12), 42.6 (C-13), 49.5 (C-14), 25.4 (C-15), 28.3 (C-16), 54.8 (C-17), 12.0 (C-18), 22.9 (C-19), 35.3 (C-20), 18.6 (C-21), 31.7 (C-22), 24.8 (C-23), 68.4 (C-24), 14.2/14.3 (CH<sub>3</sub>-CH<sub>2</sub>-O), 63.7/63.8 (CH<sub>3</sub>-CH<sub>2</sub>-O), 155.3/ 154.5 (O<sub>2</sub>C=O).

 $3\alpha$ , 24-Dihydroxy-5 $\beta$ -cholan-7-one (6): A mixture of 600 mg of 5 (1.59·10<sup>-3</sup> mol) and 10 ml of 5% methanolic KOH was stirred at room temp. for 15 min and then poured into water. The solid was collected by filtration, washed with abundant water and dried at 60°C. Chromatographic separation on silica gel with hexane/ethyl acetate (3:2) as solvent system gave 391 mg of 6 (90%), white crystalline solid, m.p. 148-149 °C. – IR (KBr):  $\tilde{v}=3496,\ 3350\ cm^{-1}$ (OH), 1713 (C=O). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 3.58$  (m, 1 H, 3- $H_{ax}$ ), 0.66 (s, 3 H, 18-H), 1.20 (s, 3 H, 19-H), 0.94 (d, J = 6.5 Hz, 3 H, 21-H), 3.58 (t, 2 H, 24-H). - <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 34.1$ (C-1), 29.6 (C-2), 70.5 (C-3), 37.1 (C-4), 42.8 (C-5), 45.4 (C-6), 213.0 (C-7), 49.5 (C-8), 46.1 (C-9), 35.1 (C-10), 21.6 (C-11), 38.9 (C-12), 42.5 (C-13), 48.8 (C-14), 24.7 (C-15), 28.3 (C-16), 54.9 (C-17), 11.9 (C-18), 23.0 (C-19), 35.4 (C-20), 18.6 (C-21), 31.8 (C-22), 29.2 (C-23), 63.1 (C-24). – MS (70 eV); m/z (%): 376 (9.2) [M<sup>+</sup>],  $358 (13.7) [M^+ - H_2O], 340 (7.8) [M^+ - 2H_2O], 299 (3.9) [M^+ - 2H_2O], 290 (3.9) [M^+ - 2H$  $H_2O - C_3H_7O$ ], 271 (7.8) [M<sup>+</sup> -  $H_2O - C_5H_{11}O$ ], 41 (100) [C=  $C-C^{+}$ ]. -  $C_{24}H_{40}O_{3}$  (376.58): calcd. C 76.55, H 10.71; found C 76.49, H 10.70.

 $3\alpha,24\text{-}Dicathyloxy\text{-}B\text{-}homo\text{-}8\text{-}oxa\text{-}5\beta\text{-}cholan\text{-}7\text{-}one}$  (7): To a solution of trifluorperoxyacetic acid [prepared from 30% aqueous  $H_2O_2$  (0.7 ml, 5.85 mmol) and TFAA (3.7 ml)] in 5 ml of chloroform, a solution of 216 mg of **6** (0.42 \cdot 10^{-3} mol) in 15 ml of chloroform was added dropwise at 0°C and the mixture was stirred for 5 min and diluted with 10 ml of chloroform. The resulting solution was washed with water (1  $\times$  3 ml), aqueous  $Na_2CO_3$  (1  $\times$  3 ml), aqueous  $NaHSO_3$  (1  $\times$  3 ml), brine (1  $\times$  3 ml) and water (1  $\times$  3 ml),

and dried with MgSO<sub>4</sub>. The solvent was removed by evaporation to give 209 mg of 5 (94%), white crystalline solid, m.p. 92.4-94.4°C. – IR (KBr):  $\tilde{v} = 1743 \text{ cm}^{-1}$  (C=O). – <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 4.59$  (m, 1 H, 3-H<sub>ax</sub>), 4.18 (t, 1 H, 8-H<sub>ax</sub>), 0.66 (s, 3 H, 18-H), 1.08 (s, 3 H, 19-H), 0.92 (d, J = 6.5 Hz, 3 H, 21-H), 4.09 (t, 2 H, 24-H), 1.29-1.31 (t, J = 7.1 Hz, 6 H,  $CH_3CH_2OCO_2$ ), 4.18 (q, J = 7.1 Hz, 4 H,  $CH_3CH_2OCO_2$ ).  $- {}^{13}C$  NMR (CDCl<sub>3</sub>):  $\delta = 35.3$  (C-1), 26.4 (C-2), 76.3 (C-3), 31.6 (C-4), 41.5 (C-5); 35.5 (C-6), 173.9 (C-7), 79.6 (C-8), 39.9 (C-9), 37.5 (C-10), 22.5 (C-11), 38.4 (C-12), 43.2 (C-13), 54.4 (C-14), 25.1 (C-15), 27.8 (C-16), 55.8 (C-17), 11.5 (C-18), 23.4 (C-19), 35.1 (C-20), 18.4 (C-21), 31.7 (C-22), 25.3 (C-23), 68.3 (C-24), 14.2/14.3 (CH<sub>3</sub>-CH<sub>2</sub>-O), 63.8  $(CH_3 - CH_2 - O)$ , 155.3/154.5  $(O_2C = O)$ .

 $3\alpha,24$ -Dihydroxy-B-homo-8-oxa-5 $\beta$ -cholan-7-one (8): A mixture of 200 mg of 7 (0.51· $10^{-3}$  mol) and 10 ml of 1% methanolic  $K_2CO_3$ was stirred at room temp. for 15 h and then poured into water. The solid was collected by filtration, washed with abundant water and dried at 60°C. Chromatographic separation on silica gel with hexane/ethyl acetate (3:2) as solvent system gave 130 mg of 8 (89%), white crystalline solid, m.p. 119.0-119.4 °C. - IR (KBr):  $\tilde{\nu}=3342$ cm<sup>-1</sup> (OH), 1743 (C=O). - <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 3.58$  (m, 1 H,  $3-H_{ax}$ ), 4.21 (t, 1 H, 8- $H_{ax}$ ), 0.67 (s, 3 H, 18-H), 1.07 (s, 3 H, 19-H), 0.93 (d, J = 6.4 Hz, 3 H, 21-H), 3.58 (t, 2 H, 24-H).  $- {}^{13}$ C NMR (CDCl<sub>3</sub>):  $\delta = 35.6$  (C-1), 29.9 (C-2), 70.4 (C-3), 35.7 (C-4), 41.6 (C-5) 35.6 (C-6) 174.9 (C-7), 80.0 (C-8), 39.8 (C-9), 37.4 (C-10), 22.4 (C-11), 38.4 (C-12), 43.1 (C-13), 54.3 (C-14), 25.0 (C-15), 27.8 (C-16), 55.9 (C-17) 11.4 (C-18), 23.4 (C-19), 35.2 (C-20), 18.5 (C-21), 31.7 (C-22), 29.3 (C-23), 63.1 (C-24). - MS (70 eV); m/z

(%): 392 (2.9) [M<sup>+</sup>], 374 (8) [M<sup>+</sup> -  $H_2O$ ], 356 (7.2) [M<sup>+</sup> -  $2H_2O$ ],  $315\ (1.5)\ [M^{+}\ -\ H_{2}O\ -\ C_{3}H_{7}O],\ 305\ (4.3)\ [M^{+}\ -\ C_{5}H_{11}O],\ 287$ (4.3)  $[M^+ - H_2O - C_5H_{11}O]$ , 41 (100)  $[C=C-C^+]$ .  $- C_{24}H_{40}O_4$ (392.58): calcd. C 73.43, H 10.27; found C 73,49, H 10.35.

[1] H. G. Cluter, T. Yokota, G. Adam (Eds.), "Brassinosteroids, Chemistry, Bioactivity and Applications", A. C. S. Symp. Ser.

1991, 474.

[2] [2a] K. Mori, M. Sakakibara, Y. Ichikawa, H. Ueda, K. Okada, W. Kondo, Tetrahedron T. Umemura, G. Yabuta, S. Kuwahara, M. Kondo, *Tetrahedron* **1982**, *38*, 2099. – [<sup>2b]</sup> S. Takatsuto, N. Ikekawa, *Tetrahedron Lett.* **1983**, *24*, 773. – [<sup>2c]</sup> T. G. Back, K. Brunner, M. V. Krishna, E. Lai, *Can. J. Chem.* **1989**, *67*, 1032.

[3a] T. C. Mc Morris, P. A. Patil, R. G. Chavez, M. E. Baker, S. D. Clouse, *Phytochemistry* **1994**, *36*, 585. – [3b] C. Brosa, J. M. Capdevila, I. Zamora, *Tetrahedron* **1996**, *52*, 2435.

V. Marquardt, G. Adam, *Phytochemistry* **1986**, *25*, 1787. – M. A. Iglesias Arteaga, R. Pérez, V. Leliebre, C. S. Pérez, F. Coll, Rev. ČENIC Cienc. Quim. 1996, 27, 3.

Coll, Rev. CENIC Cienc. Quim. 1990, 27, 3.

[5] [5a] F. Coll, E. Alonso, M. A. Iglesias Arteaga, V. Marquadt, G. Adam, Rev. Cub. Quim. 1992, 6, 7. — [5b] M. A. Iglesias Arteaga, V. Leliebre, C. S. Pérez, F. Coll. Quim. Nova 1997, 20, 361.

[6] [6a] M. A. Iglesias Arteaga., R. Pérez, V. Leliebre, C. Pérez, F. Coll, J. Chem. Res. (S) 1996, 504. — [6b] M. A. Iglesias Arteaga, R. Pérez, V. Leliebre, C. S. Pérez, F. Coll, Synth Commun., 1998, 28, 1381. — [6c] M. A. Iglesias Arteaga, R. Pérez, V. Leliebre, C. S. Pérez, F. Coll. Synth Commun. 1998, 28, 1779. . Pérez, F. Coll, Synth Commun., 1998, 28, 1779.

[7] T. Iida, T. Tamaru, F. C. Chang, J. Goto, *J Lipid Res.* 1991,

 D. Leibfritz, J. D. Roberts, J. Am. Chem. Soc. 1979, 95, 4996.
 T. C. Mc Morris, P. A. Patil, J. Org. Chem. 1993, 58, 2338.
 K. Wada, S. Marumo, H. Abe, T. Morishita, K. Nakamura, M. Ushiyama, K. Mori, Agric. Biol. Chem. 1984, 48, 719.

[98035]